

## AMENDMENTS TO THE SPECIFICATION

On page 49, beginning on line 21, last paragraph bridging pages 49 and 50, amend as follows:

Total RNA was prepared from STRO-1<sup>BRT</sup>/CD146<sup>+</sup> sorted BMMNCs, and control cells (primary BMSSC cultures grown in the presence of 10<sup>-7</sup> M dexamethasone for three weeks) using RNA STAT-60 (TEL-TEST Inc. Friendswood TX). First-strand cDNA synthesis was performed with a first-strand cDNA synthesis kit (GIBCO BRL, Life Technologies) using an oligo-dT primer. First strand cDNA (2 µl) was added to 46 µl of a 1X PCR master reaction mix (Roche Diagnostics, GmbH Mannheim Germany) and 10 pMol of each human specific primer sets: CBFA1 (632bp, and three smaller alternative splice variants)<sup>(27)</sup> sense 5'-CTATGGAGAGGACGCCACGCCTGG-3' [SEQ ID NO. 1], antisense, 5'-CATAGCCATCGTAGCCTTGTCCT-3' [SEQ ID NO. 2]; osteocalcin (310bp)<sup>(4)</sup> sense, 5'-CATGAGAGCCCTCACA-3' [SEQ ID NO. 3], antisense, 5'-AGAGCGACACCCTAGAC-3' [SEQ ID NO. 4]; GAPDH (800bp) <sup>(4)</sup> sense, 5'-AGCCGCATCTTCTTTTGCCTC-3' [SEQ ID NO. 5]; antisense 5'-TCATATTTGGCAGGTTTTTCT-3' [SEQ ID NO. 6]. The reactions were incubated in a PCR Express Hybaid thermal cycler (Hybaid, Franklin, MA) at 95°C for 2 minutes for 1 cycle then 94°C/(30 sec), 60°C/(30 sec), 72°C/(45 sec) for 35 cycles, with a final 7 minute extension at 72°C. Following amplification, each reaction was analyzed by 1.5% agarose gel electrophoresis, and visualized by ethidium bromide staining.

On page 65, Table 2, amend as follows:

**Table 2.** RT-PCR primers and conditions for the specific amplification of human mRNA

<b>Target Gene</b>	<b>Sense/ Antisense (5'-3') Primer Sequences</b>	<b>Product Size</b>
<b>GAPDH</b>	CACTGACACGTTGGCAGTGG/ [SEQ ID NO. 7] CATGGAGAAGGCTGGGGCTC [SEQ ID NO. 8]	417
<b>Leptin</b>	ATGCATTGGGAACCCTGTGC/ [SEQ ID NO. 9] GCACCCAGGGCTGAGGTCCA [SEQ ID NO. 10]	492
<b>CBFA-1</b>	GTGGACGAGGCAAGAGTTTCA/ [SEQ ID NO. 11] TGGCAGGTAGGTGTGGTAGTG [SEQ ID NO. 12]	632
<b>OCN</b>	ATGAGAGCCCTCACACTCCTC/ [SEQ ID NO. 13] CGTAGAAGCGCCGATAGGC [SEQ ID NO. 14]	289
<b>GFAP</b>	CTGTTGCCAGAGATGGAGGTT/ [SEQ ID NO. 15] TCATCGCTCAGGAGGTCCTT [SEQ ID NO. 16]	370
<b>Nestin</b>	GGCAGCGTTGGAACAGAGGTTGGA/ [SEQ ID NO. 17] CTCTAAACTGGAGTGGTCAGGGCT [SEQ ID NO. 18]	460
<b>GATA-4</b>	GACTTCTCAGAAGGCAGAG/ [SEQ ID NO. 19] CTATCCTCCAAGTCCCAGAG [SEQ ID NO. 20]	800
<b>PDGFβ-R</b>	AATGTCTCCAGCACCTTCGT/ [SEQ ID NO. 21] AGCGGATGTGGTAAGGCATA [SEQ ID NO. 22]	650
<b>Osterix</b>	GGCACAAAGAAGCCGTA CT/ [SEQ ID NO. 23] CACTGGGCAGACAGTCAGAA [SEQ ID NO. 24]	247
<b>COL X</b>	AGCCAGGGTTGCCAGGACCA/ [SEQ ID NO. 25] TTTTCCCACTCCAGGAGGGC [SEQ ID NO. 26]	387
<b>SOX9</b>	CTC TGC CTG TTT GGA CTT TGT/ [SEQ ID NO. 27] CCT TTG CTT GCC TTT TAC CTC [SEQ ID NO. 28]	598
<b>Ang-1</b>	CCAGTCAGAGGCAGTACATGCTA AGAATTGAGTTA/ [SEQ ID NO. 29] GTTTTCCATGGTTTTGTCCCGCAGTA [SEQ ID NO. 30]	300